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Charge-transfer-to-solvent reactions from I^- to water, methanol, and ethanol studied by time-resolved photoelectron spectroscopy of liquids

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The charge-transfer-to-solvent (CTTS) reactions from iodide (I^-) to H_2O , D_2O , methanol, and ethanol were studied by time-resolved photoelectron spectroscopy of liquid microjets using a magnetic bottle time-of-flight spectrometer with variable pass energy. Photoexcited iodide dissociates into a weak complex (a contact pair) of a solvated electron and an iodine atom in similar reaction times, 0.3 ps in H_2O and D_2O and 0.5 ps in methanol and ethanol, which are much shorter than their dielectric relaxation times. The results indicate that solvated electrons are formed with minimal solvent reorganization in the long-range solvent polarization field created for I^- . The photoelectron spectra for CTTS in H_2O and D_2O —measured with higher accuracy than in our previous study [Y. I. Suzuki *et al.*, Chem. Sci. **2**, 1094 (2011)]—indicate that internal conversion yields from the photoexcited I^{*-} (CTTS) state are less than 10%, while alcohols provide 2–3 times greater yields of internal conversion from I^{*-} . The overall geminate recombination yields are found to be in the order of $H_2O > D_2O > \text{methanol} > \text{ethanol}$, which is opposite to the order of the mutual diffusion rates of an iodine atom and a solvated electron. This result is consistent with the transition state theory for an adiabatic outer-sphere electron transfer process, which predicts that the recombination reaction rate has a pre-exponential factor inversely proportional to a longitudinal solvent relaxation time. Published by AIP Publishing. [<http://dx.doi.org/10.1063/1.4960385>]

I. INTRODUCTION

Charge-transfer-to-solvent (CTTS) reactions,^{1–3} which involve the transfer of an electron from a solute to its solvent, offer an excellent opportunity to study solvation dynamics in chemical reactions. CTTS reactions of atomic halogen anions are of particular interest, because these solutes have no rovibrational degrees of freedom that would otherwise complicate the dynamics. Among them, CTTS from iodide (I^-) to water has been the most extensively studied,^{4–22} as water is the most important solvent and the UV absorption band of hydrated iodide, $I^-(aq)$, is the most easily accessible; the longest-wavelength UV absorption band of $I^-(aq)$ is peaked at 225 nm (220 and 219 nm in methanol and ethanol, respectively). Although I^- in the gas phase has no stable excited state, the characteristic UV absorption bands of $I^-(aq)$ arise from metastable excited states created by solvation. We denote these metastable states as I^{*-} in this paper (they are often referred to as CTTS states in the literature). Each I^{*-} state has a singly occupied diffuse molecular orbital, composed primarily of the 6s orbital of I^- and the neutral iodine core in one of the $I(^2P_{3/2})$ or $I(^2P_{1/2})$ spin-orbit states. In the present study, we focus on the $I^{*-}(^2P_{3/2})$ state and hereafter omit the $^2P_{3/2}$ notation.

If we approximate liquid water as a wide gap semiconductor, $I^-(aq)$ is an impurity and its UV absorption

occurs within the band gap. For this system, CTTS is the adiabatic separation of an excess electron and an iodine atom to form their weak complex. The complex varies in nature due to the inhomogeneity in the initial solvation structure of I^- ; however, previous studies have illustrated an average picture of the complex, evolving from a contact pair (CP), which is an excess electron and iodine pair held in the same solvation shell with excess internal energy, to a solvent-separated state (SS), which is a weak complex between a solvated electron and an iodine atom at an equilibrium temperature.^{13,20} The CP may not be a stable species,^{6,7} but it is considered to be a transient non-stationary state with a lifetime of less than 0.5 ps. Some studies have indicated that a neutral halogen atom forms a 1:1 complex with a water molecule. For instance, Sevilla *et al.* have explained their electron spin resonance (ESR) spectrum of Cl in water by assuming a H_2O-Cl complex;²³ their calculations at the UMP2/6-31G* level suggested that the O–Cl distance is 0.263 nm. A similar 1:1 pair has also been predicted for Br¹⁶ and I.¹⁷ The predicted binding energy between iodine and water is 50 meV, and MD simulations suggest that the complex dissociates on a sub-picosecond time scale once a stronger hydrogen-bonding network is constructed around the iodine.¹⁷ The H_2O-X complex, if any, is formed after electron photodetachment from I^- , and the role of this complex in CTTS remains unclear. Thus, we will not take it into account in this study.

After I^{*-} is adiabatically separated into an excess electron and an iodine atom, they undergo thermal diffusion that

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competes with geminate recombination in the potential of mean force. These dynamics, which take place in tens of picoseconds or longer, play a decisive role in determining the overall yield of free solvated electron [$e^-(\text{solv})$]. Staib and Borgis^{6,7} have indicated that the potential of mean force in the CTTS from Cl^- to water has a free energy well of $3k_B T$, where k_B is the Boltzmann constant and T is the temperature. The minimum of this potential was predicted to occur at an electron-chlorine distance of 0.6 nm, which corresponds to a structure with one water molecule between the center of an excess electron density distribution and a chlorine atom.^{6,7} It is noted that since the hydrated electron has a delocalized electron density distribution over multiple hydration shells,^{24–26} one water molecule will not completely attenuate the exchange interactions between the excess electron and the chlorine atom. The potential of mean force becomes flat at a distance of 0.8 nm, which is sufficient for three water molecules to fit in between the excess electron and the chlorine atom. Based on the theoretical calculations of Staib and Borgis, Kloepper *et al.* have analyzed their transient absorption spectra of CTTS from I^- to water¹⁰ and suggested that the potential minimum lies at 0.4 nm and has a well depth of 620 cm^{-1} .¹⁰ In a similar study carried out with higher time resolution over a wider probe wavelength region, Iglev *et al.* proposed that the potential minimum occurs at 0.6 nm and that the depth is $850 \pm 100 \text{ cm}^{-1}$.¹⁴

Previously, we reported the first time-resolved photoelectron spectroscopy (TRPES)¹⁸ study of the CTTS reaction from I^- to bulk water using liquid microjets.²⁰ TRPES using sub-picosecond UV pulses has revealed a rapid increase in the electron binding energy (eBE) over time, corresponding to evolution from CP to a hydrated electron [$e^-(\text{aq})$] via SS.²⁰ However, the low electron detection efficiency of hemispherical electron energy analyzers has limited the signal-to-noise ratio and accuracy in measurements of low-energy electrons ($<3 \text{ eV}$). Thus, our recent TRPES studies^{27,28} on other reactions were performed using a magnetic bottle time-of-flight (MBTOF) photoelectron spectrometer²⁸ with variable pass energy, which provides several orders higher signal count rate and improved accuracy in measurements of low-energy electrons compared to a hemispherical analyzer. Thus, the present study revisits the CTTS reaction from I^- to bulk water with the superior MBTOF apparatus. The new experimental data enable refinement of our previous study on the isotope effect in CTTS reactions in water.²⁰ Moreover, we investigate similar CTTS dynamics from I^- to methanol and ethanol to gain further insight into the CTTS dynamics in polar protic solvents. Previously, Vilchiz *et al.*¹² studied CTTS from I^- to alcohols using transient absorption spectroscopy (TAS), but their study focused only on the long-time dynamics of diffusion and recombination. The present study considers the dynamics more thoroughly, beginning from ultrafast solvation and including the long-time diffusion and recombination processes. We employ both kinetic and diffusion models for the data analysis, as well as compare our results on methanol with those of a recent TRPES study by Elkins *et al.*²²

II. EXPERIMENTAL SECTION

A. Liquid microjet

We employed NaI as a solute and H_2O , D_2O , methanol, and ethanol as the solvents. The concentration of the solutions was adjusted to 40–100 mM to minimize electrokinetic charging, which occurs for liquids discharged from a fused silica capillary nozzle. The sample solution was pressurized using a high-performance liquid chromatography (HPLC) gradient flow pump and degassed using an in-line degasser prior to discharging from the capillary (15 μm i.d., 10 mm length). The outer surface of the capillary was coated with fine graphite to prevent it from charging up. The laser beams were crossed with the liquid microjet at around 1 mm downstream from the nozzle. The sample solutions and the nozzle assembly were kept at room temperature, while the liquid microjet temperature was low, owing to evaporative cooling of the jet in vacuum. When the jet diameter is small ($<10 \mu\text{m}$), evaporated gas effuses into the vacuum at the liquid temperature, while the liquid microjet is cooled, as predicted by the Clausius-Clapeyron equation.^{29,30} When the microjet diameter is larger, evaporated gas stagnates above the liquid surface, and its free expansion lowers the gas temperature to below that of the liquid surface. The liquid is less efficiently cooled than expected from the Clausius-Clapeyron equation because a fraction of the stagnated gas recondenses into the liquid. This behavior has been confirmed experimentally for liquid microjets. The temperatures of our liquid microjets were estimated to be around 278 K for water and 260 K for alcohol based on the Raman thermometry study by Wilson *et al.*³¹ At these temperatures, the mutual diffusion coefficients of a pair of iodine and $e^-(\text{solv})$ in H_2O , D_2O , methanol, and ethanol are estimated to be 3.0, 2.6, 1.6, and $0.8 \times 10^{-5} \text{ cm}^2/\text{s}$, as explained in the supplementary material.⁴⁷ We have corrected the observed photoelectron kinetic energies (PKEs) for streaming potentials using the method described by Kurahashi *et al.*³²

B. Laser system

The experiments were performed using a 1-kHz Ti:sapphire regenerative amplifier (Quantronix Titan; 100 fs, 2 mJ/pulse, 800 nm) and two collinear optical parametric amplifiers (Light Conversion TOPAS). The output of one of the optical parametric amplifiers was mixed with 800 nm to generate the pump pulses at 225 nm, while the 260-nm output from the other was used as the probe pulses. The probe pulses were optically time-delayed with respect to the pump pulses using a corner cube mounted on a computer-controlled linear translation stage. Using concave mirrors, the pump and probe pulses were gently focused on the liquid microjet, at which the laser beam diameters were estimated to be 100 μm . The typical pump and probe pulse energies were 1.5–4.0 nJ/pulse and 3.6–13.0 nJ/pulse, respectively, and the cross-correlation time between them was measured using $(1 + 1')$ two-color resonance enhanced multiphoton ionization of NO to be 250–300 fs. Based on the density of iodide ($6.4 \times 10^{-2} \text{ nm}^{-3}$) and photoexcitation efficiency

(0.1%), the density of photoexcited iodide is estimated to be $6.4 \times 10^{-5} \text{ nm}^{-3}$.

C. TRPES of liquids

Photoemission from liquids is understood using a three-step model, which assumes (1) promotion of an electron into the conduction band of solvent, (2) transport of the electron to the liquid-gas interface, and (3) ejection of the electron from the interface. The upper states reached in step (1) can be either a delocalized electronic state and/or some metastable resonance states with unknown character. In step (2), an electron wave packet travels through the solvent, in which elastic and inelastic scattering can occur with solvent and/or solute molecules.^{27,28,33,34}

Since the PKE created in our UV photoemission experiment is smaller than the electronic excitation energy of the solvents ($\sim 7 \text{ eV}$), inelastic scattering in solution is primarily vibrational.³⁵ Although the cross section of vibrational inelastic scattering is smaller than that of electronic inelastic scattering, its finite magnitude determines the effective attenuation length of low-energy electrons in solvent. Since elastic scattering deflects the velocity directions of electrons and increases the effective path length in liquid, it increases the probabilities of inelastic scattering events. Although effective attenuation lengths at low PKEs ($< 10 \text{ eV}$) have not been established, experimental studies suggest that they are on the order of nanometers.^{33,36–40}

D. Magnetic bottle spectrometer

The photoionization chamber houses a liquid beam nozzle on a 3-axis manipulator and a SmCo magnet with a conical iron cap on another 3-axis manipulator. The tip of the magnet was placed 2 mm from the photoionization point. The photoionization chamber was evacuated with a 1600 l/s turbo molecular pump and a liquid nitrogen trap employed for cryo-pumping. A typical pressure in the chamber was 10^{-4} Torr when running a liquid jet. The liquid jet traveled vertically downward in the photoionization chamber and was trapped in a vessel cooled with liquid nitrogen. An entrance skimmer ($\varnothing 0.5 \text{ mm}$) for an electron TOF analyzer was located 2 mm from the ionization point. The analyzer, which contains a solenoid coil and a $\varnothing 40 \text{ mm}$ microchannel plate (MCP) detector, was evacuated using a 2600 l/s turbo molecular pump. The electric potential of the MCP surface was kept at +300 V to ensure that the detection sensitivity was independent of the initial PKE. A grounded wire mesh was placed in front of the MCP to prevent leakage of the electric field due to the positive voltage into the field-free region. The signal from the MCP was amplified using a preamplifier (FAST ComTec, TA2000B-3) and counted with a multichannel scaler (FAST ComTec, P7887). The electron flight path from the ionization point to the detector was 1090 mm. The magnetic flux densities at the ionization point and in the flight tube were estimated to be 0.4 T and 1 mT, respectively. Both the photoionization chamber and TOF analyzer were shielded against external magnetic fields by a permalloy inner layer.

The electron detection efficiency of a standard MBTOF spectrometer diminishes for low PKEs of $< 0.5 \text{ eV}$. In this study, we have employed an MBTOF spectrometer with a variable electric potential in the electron flight region to increase the kinetic energies of photoelectrons in the analyzer. Prior to the measurements, we have confirmed that the electron transmission curve of our MBTOF spectrometer was flat for PKEs higher than 0.3 eV. Therefore, we applied a voltage of +0.5 V to the analyzer to make the electron pass energy through the analyzer higher than 0.5 eV, which ensured that the detection efficiency was uniform over the entire PKE region. For a 0.1 M aqueous NaI solution, an average number of ~ 2.5 electrons were detected per laser shot at pump and probe pulse energies of 3 and 13 nJ/pulse, respectively.

III. MODELS FOR DATA ANALYSIS

In this study, we employed both kinetic and diffusion models for analyzing our TRPES data and examined the consistency between them. Although the kinetic model is easily implemented, it does not describe the long-time diffusion rigorously. The diffusion model enables systematic interpretation of the observed time profiles through use of the diffusion constants. In this section, the two models are briefly explained.

A. Kinetic model

We have performed a global fit of the time-dependent electron binding energy distribution (eBED) by assuming that each transient species has a time-independent photoelectron spectrum and a time-dependent population. As we reported previously,²⁰ the observed photoelectron intensity profile exhibits at least three exponentially decaying components. Thus, we assumed two reaction intermediates, CP and SS, in our kinetic model, as shown in Fig. 1.

With this kinetic model, the relative population in each state after photoexcitation to I^{*-} is expressed as follows:

$$[\text{I}^{*-}] = e^{-t/\tau_a}, \quad (1)$$

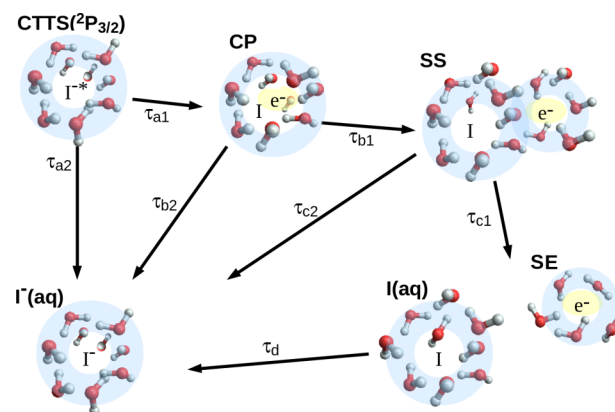


FIG. 1. Graphical representation of our kinetic model for the CTTS reaction from I^- to bulk water. τ_d in the model corresponds to the overall depopulation time of the solvated electron (SE), during which geminate recombination is dominant.

$$[\text{CP}] = r_a \tau_b \left(\frac{e^{-t/\tau_a}}{\tau_{ab}} + \frac{e^{-t/\tau_b}}{\tau_{ba}} \right), \quad (2)$$

$$[\text{SS}] = r_a r_b \tau_c \left(\frac{\tau_a e^{-t/\tau_a}}{\tau_{ab} \tau_{ac}} + \frac{\tau_b e^{-t/\tau_b}}{\tau_{ba} \tau_{bc}} + \frac{\tau_c e^{-t/\tau_c}}{\tau_{ca} \tau_{cb}} \right), \quad (3)$$

$$[\text{e}^- (\text{solv})] = r_a r_b r_c \tau_d \left(\frac{\tau_a^2 e^{-t/\tau_a}}{\tau_{ab} \tau_{ac} \tau_{ad}} + \frac{\tau_b^2 e^{-t/\tau_b}}{\tau_{ba} \tau_{bc} \tau_{bd}} + \frac{\tau_c^2 e^{-t/\tau_c}}{\tau_{ca} \tau_{cb} \tau_{cd}} + \frac{\tau_d^2 e^{-t/\tau_d}}{\tau_{da} \tau_{db} \tau_{dc}} \right), \quad (4)$$

where $\tau_x = (\tau_{x1}^{-1} + \tau_{x2}^{-1})^{-1}$ and $\tau_{xy} = \tau_x - \tau_y$ ($x, y = a, b, c$, and d). The branching ratio in each kinetic step is denoted by $r_x (= \tau_x \tau_{x1}^{-1})$. The recombination time constants τ_{x2} are obtained from the fitting parameters τ_x and r_x as $\tau_{x2} = \tau_x (1 - r_x)^{-1}$. If all transient species have an identical photodetachment cross section σ_i ,²⁰ the photoelectron signal intensity is expressed by

$$I(t, E_k) = \sigma_i (c_{ak} [\text{I}^*] + c_{bk} [\text{CP}] + c_{ck} [\text{SS}] + c_{dk} [\text{e}^- (\text{solv})]) \otimes g(t), \quad (5)$$

where k is an index for equally spaced PKE bins (typically 0.2 eV). The expansion coefficients c_{xk} ($x = a, b, c, d$) provide the eBED intensities of each chemical species in the k th energy bin. The function $g(t)$ is a Gaussian cross-correlation function representing our laser pulses. It is possible that the actual photodetachment cross sections of each species differ; however, they are not separated mathematically from the r_x values and cannot be rigorously determined. Thus, the validity of the assumed identical cross sections is discussed later in combination with the r_x values extracted from the data analysis.

B. Diffusion model

The photoelectron signal intensity at long times (>several ps) following photoexcitation of I^- is primarily determined by mutual diffusion and recombination of a solvated electron and an iodine atom, which can be analyzed using the Smoluchowski diffusion equation. On the other hand, the initial reaction step from I^* to CP (or SS) is not a diffusion process, and it should be described as a kinetic step(s). As the minimal assumption necessary for our diffusion model, we employed only an exponential decay of I^* with time constant τ_a to form a weak complex; the complex is generated at an electron-iodine distance of r_0 in the potential of mean force with time-dependence $r_a \{1 - \exp(-t/\tau_a)\}$, where r_a is the quantum yield of the complex. As we show later, least squares fitting without assuming a kinetic step can also reproduce the observed time profile; however, it requires an unrealistic potential of mean force. In our diffusion model analysis, no photoelectron spectral evolution is taken into account; in fact, the spectral evolution is negligible after formation of a weak complex of an iodine atom and an electron.

The probability density, $c(t, r)$, of a solvated electron around an iodine atom depends on the elapsed time (t) after irradiation by a pump pulse and the iodine-electron distance (r). We assume spherical symmetry for the probability

density⁴¹ and a sufficiently low concentration of I^* so that only a single electron-iodine pair in the system can be considered. The diffusion equation is written as

$$\frac{\partial \psi(t, r)}{\partial t} = \hat{L} \psi(t, r), \quad (6)$$

where $\psi(t, r)$ and \hat{L} are, respectively,

$$\psi(t, r) = 4\pi r^2 c(t, r) \quad (7)$$

and

$$\hat{L} = D' \frac{\partial}{\partial r} r^2 e^{-V(r)} \frac{\partial}{\partial r} r^{-2} e^{V(r)}. \quad (8)$$

In the above, $V(r)$ is the potential of mean force (in units of $k_B T$), and D' is the mutual diffusion constant.

Equation (6) is solved numerically by discretizing the radial coordinate r with an equal distance Δr of 5 pm. When expressing $\psi(t, r)$ in each cell as $q_i(t)$ ($i = 1, 2, \dots, N$), $q_i(t)$ obeys the following equation:

$$\begin{aligned} \frac{d}{dt} q_i(t) = & \Omega(i|i-1) q_{i-1}(t) - [\Omega(i-1|i) + \Omega(i+1|i)] \\ & \times q_i(t) + \Omega(i|i+1) q_{i+1}(t), \end{aligned} \quad (9)$$

where $\Omega(i|j)$ is the transition probability from j to i per unit time. Equation (9) is equivalently expressed in vector-matrix form as

$$\frac{d}{dt} \mathbf{q} = \mathbf{L} \cdot \mathbf{q}. \quad (10)$$

We assume that geminate recombination occurs at the contact radius r_{contact} ($r_{\text{contact}} < r_0$). As a boundary condition, geminate recombination is introduced at r_{contact} as $\Omega(0|1) = \kappa_r / \Delta r$. Equation (10) is solved by diagonalizing the matrix \mathbf{L} , and the solution is obtained in the following form:

$$q_i(t) = \sum_{j=1}^N a_{ij} \exp\left(-\frac{t}{\tau_j}\right), \quad (11)$$

where a_{ij} are the constants that depend on the initial conditions, and $-1/\tau_j$ are the eigenvalues of \mathbf{L} . Then, the total photoelectron intensity is expressed by

$$I(t) = \sigma_i \left[e^{-t/\tau_a} + \sum_{i=1}^N q_i(t) \right] \otimes g(t), \quad (12)$$

where $g(t)$ is the Gaussian cross-correlation function between the pump and probe pulses. Here, we have assumed the photodetachment cross section σ_i to be independent of r .

We have approximated the attractive potential of mean force using the Morse function,

$$V(r) = \Delta G \left(1 - e^{-\beta(r-r_{\text{min}})} \right)^2 - \Delta G, \quad (13)$$

with the potential minimum r_{min} taken to be the initial distance r_0 . Previously, Kloepper *et al.* determined a value of $\kappa_r = 0.5$ nm/ps, assuming $r_0 = r_{\text{contact}} = 0.4$ nm,¹⁰ while Iglev *et al.* determined $r_{\text{contact}} = 0.5 \pm 0.01$ nm, assuming $r_0 = 0.61$ nm and $\kappa_r = \infty$ (the absorbing boundary condition).¹⁴ In our analysis, we also assumed a value of $r_0 = r_{\text{contact}} = 0.4$ nm to determine the five parameters τ_a , r_a , ΔG , β , and κ_r . Iglev *et al.*¹⁴ argued that the value of $r_{\text{contact}} = 0.4$ nm assumed by Kloepper *et al.*¹⁰ is too small and attributed it to be the source of their shallower interaction

potential; however, this argument is not quite correct. In fact, a smaller assumed r_0 and r_{contact} should provide a deeper interaction potential, so the shallower potential estimated by Klopfer *et al.*¹⁰ should be due to other factors. Since it was difficult for us to accurately determine the absolute values of r_0 and r_{contact} , we assumed $r_0 = r_{\text{contact}} = 0.4$ nm in all of our analyses; however, this simplification does not qualitatively alter our conclusion.

IV. RESULTS AND ANALYSIS

A. CTTS in water and kinetic model analysis

Figure 2 shows (a) the two-dimensional (2D) map of the eBED measured for CTTS in H₂O, (b) the global fit using the kinetic model, and (c) the residue of the fit. The time axis is logarithmic. The global fit well reproduces the experimental 2D map with the best-fit parameters listed in Table I. The overall decay time constants, τ_a , τ_b , and τ_c , are in agreement with our previous study and also with the time constants of 0.22 ± 0.05 , 0.7 ± 0.05 , and 21 ± 2 ps reported by Iglev *et al.* using TAS for a 2.5 mM NaI solution at 298 K.¹⁴ It is worth noting that depopulation of the I^{-*} state may not occur immediately after photoexcitation and that the signal exhibits a plateau. Careful analysis of the results shown in Fig. 4 and similar data obtained with a higher time resolution of 125 fs indicates that the experimental results can also be explained by assuming an induction time of about 50 fs prior to I^{-*} population decay and a decay time constant (τ_a) of ~ 100 fs. This fact seems to accord with the 60 fs component observed by Chergui and colleagues using ultrafast fluorescence spectroscopy of this reaction;²¹ however, the time resolution of the present study is not quite sufficient to elucidate the sub-100 fs dynamics clearly.

Figure 4 shows the decay-associated spectra determined for the four solvents using the global fit. The spectral intensities of CP, SS, and e⁻(solv) in H₂O and D₂O appear stronger than those of our previous study due to the improved detection sensitivity for low-energy electrons and

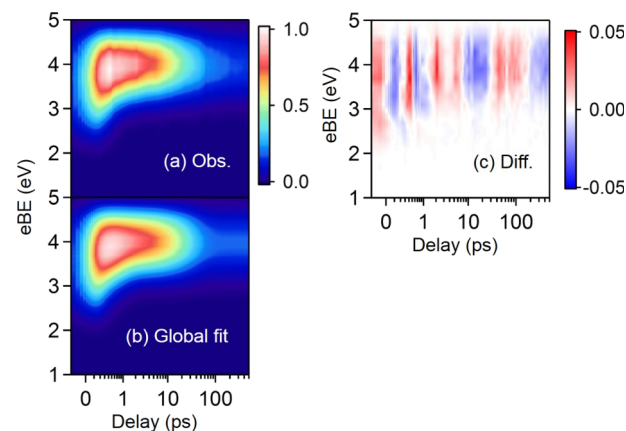


FIG. 2. (a) Time-evolution of the eBED measured for the CTTS reaction from I^{-*} to H₂O, (b) simulated distribution obtained by least squares fitting based on the kinetic model, and (c) the difference between (a) and (b). The pump and probe wavelengths are 225 and 260 nm, respectively. A constant has been added to the actual delay ($t_{\text{plot}} = t_{\text{true}} + 0.2$ ps) to shift the entire distribution and display the data around $t = 0$. The time label and grids are presented for t_{true} .

accurate calibration of the electron transmission curve of the electron spectrometer. The integrated area of each spectrum in Fig. 3 provides the quantum yield multiplied by the relative photodetachment cross section of each species. By assuming the photodetachment cross sections of I^{-*} and CP to be identical, the quantum yield (r_a) of CP is estimated to be 0.95 in H₂O and 0.89 in D₂O, as listed in Table I. These values are greater than those of 0.60 and 0.54 estimated in our previous study.²⁰ This discrepancy is ascribed to the low detection efficiency of low-energy electrons and incomplete calibration of the hemispherical electron analyzer in our previous study.

If the true quantum yield of CP is unity, the smaller electron signal intensity from CP implies that the detachment cross section of CP is lower than that of I^{-*}. In either cases, this study clearly shows that the yield of CP is greater than 0.9 and that quenching (internal conversion) of I^{-*} is not significant. CTTS in H₂O and D₂O exhibits almost indistinguishable eBEDs, as can be seen in Fig. 3, as well as very similar I^{-*}

TABLE I. Kinetic time constants (ps) and branching ratios determined for each sample.

	NaI/H ₂ O	NaI/H ₂ O ^a	NaI/D ₂ O	NaI/D ₂ O ^a	NaI/MeOH	NaI/MeOH ^b	NaI/EtOH
τ_a	0.23 ± 0.03	0.23 ± 0.1	0.25 ± 0.01	0.31 ± 0.1	0.45 ± 0.09	0.55 ± 0.11	0.44 ± 0.10
τ_b	0.87 ± 0.29	1.1 ± 0.2	1.1 ± 0.2	1.33 ± 0.2	7.4 ± 1.3	8 ± 3	3.7 ± 1.1
τ_c	23 ± 2.0	21 ± 2	37 ± 1.7	37 ± 4	65 ± 9.5	41 ± 7	119 ± 25
τ_d	2.1×10^9 ^c	300–1600	2.6×10^9 ^c	400–1400	1.1×10^8 ^c		2.6×10^{15} ^c
τ_{a1}	0.25 ± 0.04	0.4 ± 0.1	0.29 ± 0.03	0.6 ± 0.1	0.64 ± 0.14	0.55	0.55 ± 0.13
τ_{a2}	4.1 ± 5.7	0.6 ± 0.1	2.1 ± 1.2	0.7 ± 0.1	1.6 ± 0.47		2.1 ± 0.57
τ_{b1}	1.3 ± 0.4	1.7 ± 0.3	1.7 ± 0.3	2.5 ± 0.3	8.2 ± 1.4	7.7	3.7 ± 1.1
τ_{b2}	2.6 ± 0.9	3.5 ± 0.5	3.1 ± 0.6	2.9 ± 0.4	73 ± 18.5		1171 ± 3240
τ_{c1}	105 ± 12	84 ± 10	135 ± 11	128 ± 20	115 ± 17.5	57	162 ± 37
τ_{c2}	29 ± 3	28 ± 2	50 ± 3	52 ± 5	147 ± 23	146	452 ± 138
r_a	0.94 ± 0.08	0.60 ± 0.05	0.88 ± 0.07	0.54 ± 0.04	0.72 ± 0.06	1	0.79 ± 0.03
r_b	0.66 ± 0.02	0.67 ± 0.01	0.64 ± 0.03	0.53 ± 0.01	0.90 ± 0.02	1	1.00 ± 0.01
r_c	0.21 ± 0.02	0.25 ± 0.01	0.27 ± 0.02	0.29 ± 0.02	0.56 ± 0.02	0.71	0.74 ± 0.06

^aReference 20.

^bReference 22.

^cThese values were too large to determine accurately in our measurements.

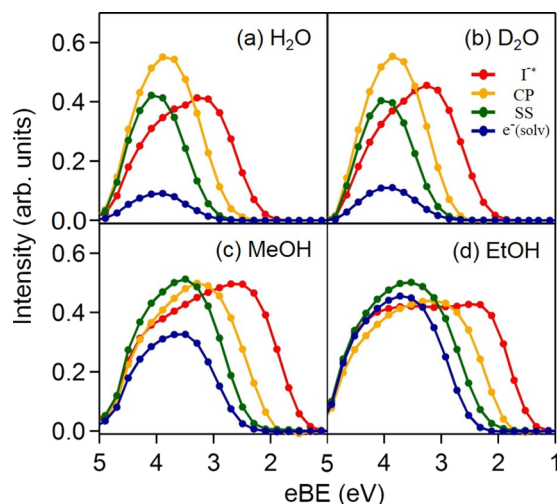


FIG. 3. Decay-associated spectra determined for CTTS reactions from I^* to (a) H_2O , (b) D_2O , (c) methanol, and (d) ethanol. The red, yellow, green, and blue curves correspond to I^* , CP, SS, and $e^-(solv)$. The pump and probe wavelengths are 225 and 260 nm, respectively.

lifetimes of $\tau_a = 0.25$ ps and 0.24 ps (Table I), respectively. The quantum yields, r_b and r_c , and associated time constants τ_{b1} , τ_{b2} , τ_{c1} , and τ_{c2} are in excellent agreement with our previous study.

B. CTTS in alcohol and kinetic model

We performed similar experiments on the CTTS reactions from I^- to methanol and ethanol. The time-energy maps of eBED presented in Fig. 4 clearly indicate that alcohol solutions exhibit higher quantum yields of $e^-(solv)$. The global fits (not shown here) using the kinetic model well reproduced the observed eBEDs. The best-fit time constants and branching ratios are tabulated in Table I. Our spectra measured for methanol exhibit greater bandwidth than those reported by Elkins *et al.*,²² which suggests that the

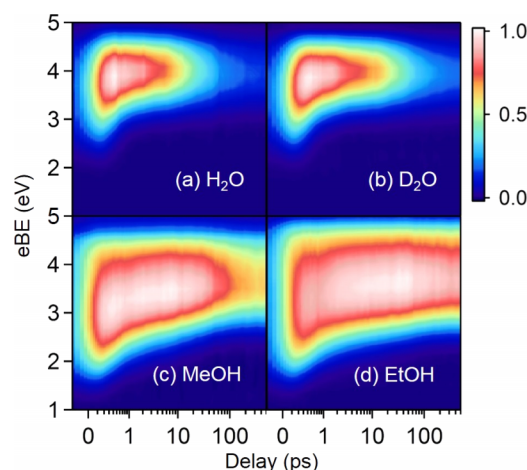


FIG. 4. Time-evolution of the eBED measured for the CTTS reaction from I^* to (a) H_2O , (b) D_2O , (c) methanol, and (d) ethanol. The pump and probe wavelengths are 225 and 260 nm, respectively. A constant has been added to the actual delay ($t_{plot} = t_{true} + 0.2$ ps) to shift the entire distribution and display the data around $t = 0$. The time label and grids are presented for t_{true} .

electron transmission efficiency of their magnetic bottle spectrometer diminishes at low electron kinetic energy. Nonetheless, the extracted time constants are in reasonable agreement between the two studies.

Although the reaction times from I^* to CP are slightly longer than that in water, they are still less than 1 ps. Interestingly, the reaction time (τ_{a1}) in ethanol is smaller than that in methanol. The r_a (0.7–0.8) values for the alcohols are smaller than that in water, while the overall quantum yields ($\phi_{SE} = r_a r_b r_c$) of $e^-(solv)$ are far greater, which is primarily due to the larger r_c values. If we neglect r_a and r_b , the survival probability of $e^-(solv)$ is given by r_c to be 0.56 and 0.74 for methanol and ethanol, respectively. These values are smaller than the corresponding values of 0.70 and 0.85 previously reported by Vilchiz *et al.* using TAS of the solutions at room temperature.¹² The differences are most likely the result of the lower temperature of our liquid microjet (estimated to be 260 K).

The decay-associated spectra extracted for methanol and ethanol are shown in Fig. 3. The photoelectron spectra for the CTTS reactions in alcohol are much broader than that in water. Specifically, the solvated electron bandwidth increases in the order of water (1.3) < methanol (1.6) < ethanol (1.9 eV) at the probe wavelength of 260 nm ($h\nu_{probe} = 4.8$ eV).²⁸ The best estimates for the vertical electron binding energies (VBES) of solvated electrons in water, methanol, and ethanol have been reported to be 3.3, 3.1, and 3.1 eV, respectively,^{18,28,42–46} while Fig. 4 indicates higher VBE values. The photoelectron spectra of I^* are rather broad in all cases, which is due to inelastic scattering in solution, as discussed in detail elsewhere.²⁸

C. Diffusion model analysis

Figures 5 and 6 show a comparison of the observed photoelectron time profile and the diffusion model simulation for methanol and ethanol, respectively. The best-fit parameters employed are summarized in Table II. We assumed $r_a = r_{min} = 0.4$ nm in all cases. The well depths ΔG of the potential of mean force estimated for methanol and ethanol are smaller than those reported by Vilchiz *et al.*¹² The discrepancy is

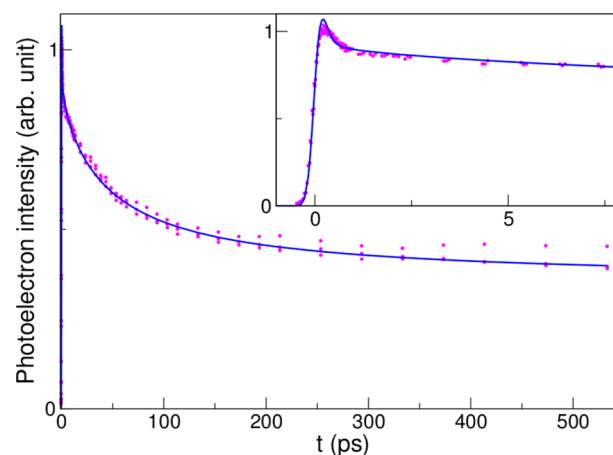


FIG. 5. The total photoelectron intensity as a function of time observed for NaI solution in methanol. The experimental data points are plotted with dots, and the solid line is the result of the least-squares fit to a diffusion model.

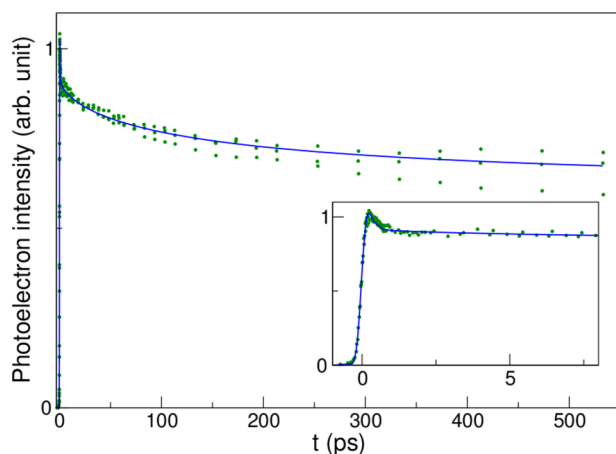


FIG. 6. The total photoelectron intensity as a function of time for NaI solution in ethanol. The experimental data points are plotted with dots, and the solid line is the result of the least-squares fit to a diffusion model.

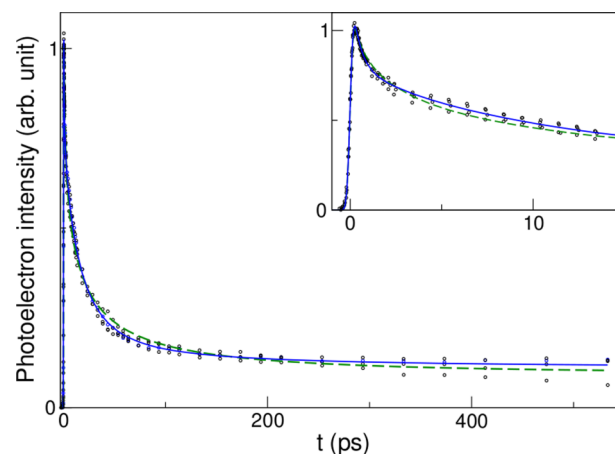


FIG. 7. The total photoelectron intensity as a function of time for aqueous NaI solution. The experimental data points are plotted with dots. The solid line is the result of the least-squares fit using a diffusion model with a metastable I^* , while the dashed line is the result obtained without assuming any metastable state. See the text for the assumed parameters.

attributed to the considerably larger diffusion coefficients that they assumed, which provides a deeper well for a given time profile. The mutual diffusion coefficients we employed for our simulation are listed in Table II and explained in the supplementary material.⁴⁷

We found it possible to fit the observed time profile of the aqueous solution reasonably well without assuming any lifetime for I^* , if an interaction potential $V(r)$ is adjusted (Fig. 7). However, such fitting provides too large of a potential width β^{-1} of about 0.3 nm. On the other hand, the diffusion model analysis assuming a finite lifetime of I^* (Table I) provides a β^{-1} of 0.11 nm and a time constant τ_a of 0.38 ps. This result confirms that CTTS from $I^-(\text{solv})$ occurs adiabatically with a finite I^* lifetime.

V. DISCUSSION

A. Ultrafast CTTS dynamics

While the shortest time constant τ_a (0.25 ± 0.01) determined for D_2O agrees with our previous value (0.31 ± 0.1) within the experimental errors,²⁰ the new value is smaller than

TABLE II. Results obtained using a diffusion model.

	H ₂ O	MeOH	EtOH	H ₂ O ^a	H ₂ O ^b	MeOH ^c	EtOH ^c
T (K)	278 ^d	260 ^d	260 ^d	297	298	297	297
$\Delta G/k_B T$	2.4	1.4	0.92	3.0	4.1	3.6	3.1
β (nm ⁻¹)	9.6	11	10	11	6.8	(10) ^e	(10) ^e
$D' \times 10^5$ (cm ² /s)	3.0 ^e	1.6 ^e	0.78 ^e	8 ^e	5.8 ^e	13.1 ^e	6.6 ^e
r_0 (nm)	4.0 ^e	4.0 ^e	4.0 ^e	4.0 ^e	6.1	4.0 ^e	4.0 ^e
τ_a (ps)	0.38	0.13	0.13	0.2	0.22
τ_b (ps)	0.7
r_a	0.70	0.52	0.57				
κ_r (pm/ps)	8.1	2.3	0.55	5.2			

^aReference 10.

^bReference 14.

^cReference 12.

^dEstimated temperature.

^eFixed values during fitting.

the previous one. Consequently, the new τ_{ai} ($i = 1$ or 2) values for H_2O and D_2O become almost indistinguishable, which indicates that the CTTS dynamics in this time range have a negligible deuterium isotope effect and that the translation of water molecules plays an important role. It is possible that the librational response of water molecules plays a role at even shorter times (<50 fs); however, this could not be confirmed in the present study due to our limited time resolution (0.3 ps). More importantly, the present study shows that the τ_{a1} values in methanol and ethanol are considerably shorter than their longitudinal solvent relaxation times, indicating that the initial reaction step from I^* to CP does not require significant reorientation of the solvent molecules.

As discussed earlier, estimation of the quantum yields of each species depends on the assumption of photodetachment cross sections of I^* , CP, SS, and $e^-(\text{solv})$. The photodetachment cross section is essentially the same for H_2O and D_2O within the Born-Oppenheimer approximation; therefore, the different r_a values extracted for H_2O and D_2O in our previous study²⁰ suggested that internal conversion takes place from the I^* state. However, in the present study, we found similarly high r_a values (0.9) for H_2O and D_2O , which cannot be regarded as strong evidence for internal conversion from I^* in water. On the other hand, the r_a values in alcohols are lower than that in water. Based on the similarities in the UV absorption spectra of I^- and the eBE of solvated electrons between water and alcohols, it is unlikely that the photodetachment cross sections differ significantly between water and alcohols. Therefore, we believe that internal conversion occurs from I^* at least in alcohol, which can be examined by observing the ground state bleach recovery of I^- .

B. Recombination and diffusion dynamics at thermal equilibrium

The mutual diffusion coefficients (D') for a pair of $e^-(\text{solv})$ and iodine under our experimental conditions are

estimated to be 3.1, 2.5, 1.7, and 0.8×10^{-5} cm²/s for H₂O, D₂O, methanol, and ethanol, respectively. Vilchiz *et al.* have argued that their rate constant k_d , which corresponds to $(\tau_{c1})^{-1}$ in our kinetic model, is proportional to the estimated D' value.¹² We found that $(\tau_{c1})^{-1}$ gradually decreases with D' ; however, we did not find a linear relationship between $(\tau_{c1})^{-1}$ and D' . This difference between the two studies is attributed to the D' values estimated inaccurately by Vilchiz *et al.* based on the viscosities of the solvents.¹² We note that the diffusion coefficient of e⁻(solv) does not scale with the viscosities of the solvents; for example, D' is larger in H₂O than in methanol, even though the former is more viscous than the latter.

Using the diffusion model and estimated D' values listed above, we found that the depth (ΔG) of the attractive potential of mean force diminishes in the order of H₂O > methanol > ethanol. We have previously estimated that the charge-induced dipole interaction between a solvated electron and neutral iodine atom is rather weak.²⁰ Thus, the main source of the potential of mean force is considered to be the hydrogen-bonding network, and the observed well depth appears to scale with the number of hydrogen bonds created per unit volume. Considering the molecular size of alcohols, the value of $r_{\min} = 0.4$ nm assumed in the diffusion analysis may be too small. However, as we mentioned earlier, assumption of a larger r_{\min} value provides a smaller well depth for alcohols, so the observed trend is unaltered by assuming longer r_{\min} values for alcohols.

One of the clearest isotope effects observed in our previous study²⁰ was for τ_{c2} , and we speculated that it originates in part from the zero point energy difference between liquid H₂O and D₂O. However, the comparison with alcohols in the present study enables a more systematic consideration of this problem. Here, we find that the quantum yield of e⁻(solv) and the diffusion coefficient are anti-correlated: the slower the diffusion, the smaller the yield of recombination. This counter-intuitive result indicates that the recombination time constant τ_{c2} differs greatly among the solvents. In fact, our kinetic model provides τ_{c2} values of 30, 150, and 450 ps for water, methanol, and ethanol, respectively, in good agreement with the values of 29, 220, and 830 ps obtained by Vilchiz *et al.*¹² The results are also consistent with the diffusion model, which yields respective κ_r values of 8.1, 2.3, and 0.55 pm/ps for water, methanol, and ethanol. Zusman has shown that an adiabatic electron transfer reaction rate is approximated by⁴⁸

$$k_{ET} = \frac{1}{\tau_L} \sqrt{\frac{\lambda}{16\pi k_B T}} \exp \left[-\frac{(\Delta G^0 + \lambda)^2}{4\lambda k_B T} \right], \quad (14)$$

where τ_L is the solvent longitudinal relaxation time given by $(\epsilon_\infty/\epsilon_0)\tau_D$, in which ϵ_∞ and ϵ_0 are, respectively, the dielectric constants at high and low frequency limits, and τ_D is the dielectric relaxation time of the solvent. ΔG^0 is the free energy of reaction, and λ is the reorganization energy. The value of λ in a dielectric continuum may be approximated as

$$\lambda = (\Delta e)^2 \left\{ \frac{1}{2a_1} + \frac{1}{2a_2} - \frac{1}{R} \right\} \left\{ \frac{1}{\epsilon_\infty} - \frac{1}{\epsilon_0} \right\}, \quad (15)$$

where Δe , a_1 , a_2 , and R are the amount of transferred charge, the radii of the donor and acceptor, and the distance between their centers. The last term $\left\{ \frac{1}{\epsilon_\infty} - \frac{1}{\epsilon_0} \right\}$ slightly diminishes from 0.55 for water to 0.50 for ethanol; however, this difference is small, because the electronic responses of these solvents are similarly fast. Soft X-ray photoemission spectroscopy shows that the eBEs of I⁻ in water and methanol are 8.0 and 7.4 eV, respectively. In addition, the eBEs of e⁻(solv) in water and methanol are estimated to differ by less than 0.2 eV, although their absolute values may still contain ambiguities.²⁸ Thus, although ΔG^0 may be slightly larger for water than for alcohols, $\Delta G^0 + \lambda$ is expected to be similar. If that is the case, k_{ET} will scale crudely with $1/\tau_L$. In fact, the ratio of τ_{c2} values for H₂O, methanol, and ethanol, 30:150:450, is similar to the ratio of τ_L values, 0.2:3:10. Our diffusion model provides the same trend for the ratio of $1/\kappa_r$ values, 0.12:0.43:1.8.

As we described in our previous study,²⁰ the quantum yield of e⁻(solv) diminishes at lower temperatures (Fig. 7 of Ref. 20). It indicates that the temperature dependence of geminate recombination is weaker than that of diffusion; therefore, the activation energy for recombination is small. Thus, the pre-exponential factor of Equation (14), which is proportional to $1/\tau_L$, becomes important for the recombination rate constants.

VI. CONCLUSION

The charge-transfer-to-solvent reactions from I⁻ to three polar protic solvents, water, methanol, and ethanol, were studied by time-resolved photoelectron spectroscopy of liquid microjets using a magnetic bottle time-of-flight spectrometer with variable pass energy. The results can be summarized as follows:

1. The elementary reaction step from I^{-*} to a contact pair occurs in less than 0.3 ps in water and 0.5 ps in alcohols. The sub-picosecond dynamics in H₂O and D₂O are quite similar, indicating that they are dominated by translational motion. The librational response of water was not detectable with our time resolution (0.3 ps). The fact that CP is formed on the sub-picosecond time scale in ethanol indicates that it is formed without significant solvent reorganization in the long-range polarization field preexisting for I⁻. The quantum yield of a contact pair is >0.9 in water and 0.7–0.8 in alcohols, which suggests that a faster solvent response time is essential to prevent internal conversion.
2. The overall quantum yield of solvated electrons is predominantly determined by geminate recombination/diffusion dynamics. The yield diminishes in the order of ethanol > methanol > water, because the geminate recombination rate increases in the order of ethanol < methanol < water. The relative recombination rates in the four solvents are correlated with the longitudinal relaxation times of the solvents, as predicted by transition state theory for an adiabatic outer-sphere electron transfer process.

3. The diffusion model analysis indicates that the potential of mean force for an electron and iodine has a well depth of $2.4k_B T$, $1.4k_B T$, and $0.92k_B T$ in water, methanol, and ethanol, respectively.

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